In-Situ TEM of Electropolymerization: Direct Monitoring of Liquid-Solid Transitions

David C. Martin

Associate Dean for Research and Entrepreneurship College of Engineering Karl W. and Renate Böer Professor Materials Science and Engineering The University of Delaware milty@udel.edu www.mseg.udel.edu cubic.mseg.udel.edu udel.edu/~milty UD-UC-UM IUCRC Planning Meeting May 14th, 2020

Organic Electronics and Electroactive Biomaterials



MRS Communications (2015), *5*, 131–153 © Materials Research Society, 2015 doi:10.1557/mrc.2015.17



Polymers/Soft Matter Prospective Article

Molecular design, synthesis, and characterization of conjugated polymers for interfacing electronic biomedical devices with living tissue

David C. Martin, Materials Science and Engineering, Biomedical Engineering, The University of Delaware, 201 DuPont Hall, Newark, Delaware 19716, USA Address all correspondence to David C. Martin at milty@udel.edu

(Received 19 February 2015; accepted 26 March 2015)

Abstract

Conjugated polymers are being considered for use at the interface between hard inorganic metallic and semiconducting electrodes and soft biological tissues. These organic materials have properties that are intermediate to these two extremes, and their chemistry, structure, and performance can be precisely manipulated over a large range. Examples of current interest included copolymers of poly(3,4-ethylene dioxy-thiophene) and poly(3,4-propylene dioxythiophene). This paper will review past efforts, recent activities, and future possibilities in this rapidly expanding area of materials research and technology.

MRS Communications, 5(2), 131-153, (2015).

http://dx.doi.org/10.1557/mrc.2015.17

Mag = 2.67 K X 1 µm	WD = 4.9 mm	EHT = 5.00 kV	Signal A = InLens	ESB Grid = 500 V	Date :20 Oct 2011	Time :16:36:18
Auriga 60-39-95 🛛 🖳	FIB Imaging = SEM	Noise Reductio	n = Line Int. Done	FIB Probe = 30KV:50pA	System Vacuum =	1.38e-006 mbar

Bionic devices: solid electronic wire - wet ionic tissue interfaces



Electrochemical Polymerization of Conjugated Polymers: PEDOT

- Controlled deposition directly onto electrode from aqueous solution (~1 V)
- Ability to use biologically active molecules as counterions
- Creates rough, soft surface that facilitates charge transport, promotes favorable biological interactions
- EDOT monomer, chemically oxidized PEDOT/PSS suspension available commercially
- Much more chemically stable than PPy: only 2 reactive hydrogens on conjugated ring

Mag = 2.67 K X 1 Auriga 60-39-95 ├ WD = 4.9 mm FIB Imaging = SE



PEDOT Molecular Design Features



Conjugated backbone: electrical and optical activity Only 2 reactive hydrogens on EDOT monomer: no branching defects No hydrogens directly on PEDOT polymer backbone: chemical stability Oxygens pendant to the backbone: enhanced electron density

Mag = 2.67 K X 1 Auriga 60-39-95

Molecular Wires



Conjugated polymers facilitate electron/hole transport along the backbone, as well as anion and cation transport via charge exchange with dopants

em Vacuum = 1.38e-006 mbar

Electrochemical Polymerization



https://ww2.chemistry.gatech.edu/reynolds/research/electrochemistry

Auriga 60-39-95

EHT = 5.00 kVNoise Reduction = Line Int. Done FIB Probe = 30KV:50pA

Electrochemical Polymerization of Conjugated Polymers: PEDOT



Electrochemical Polymerization of Conjugated Polymers: PEDOT









Zeiss Auriga 60 FIB/SEM

nm resolution
 STEM, EDS, EsB detectors
 Cryo-stage
 Omni-probe for TEM prep
 3D reconstructions
 Correlative Optical Microscopy:
 Computerized sample stage





			LEI 1.04 22:00 WD 7.4mm
PEDOT			PEDOT
Silicon-based substrate		S	ilicon-based substrate
$\begin{array}{c c c c c c c c c c c c c c c c c c c $	um HV NanoLab 2.00 k HT = 5.00 kV Signal A = Noise Reduction = Line Int.	HFW det mode WD V 2.56 µm TLD Custom 2.3 mm InLens ESB Grid = 500 V Done OLOPINA LOW:104	1 μm — Helios NanoLab Date :20 Oct 2011 Time :16:36:18 Μ ዋδο νοι μη Η 1.38e-006 mbar

PEDOT electrochemical deposition



In-situ TEM imaging of the electrochemical deposition of PEDOT – experimental setup

Macro Letters, (2015), http://dx.doi.org/10.1021/acsmacrolett.5b00479



Microfabricated Electrochemical chip



SEM images of the top electrochemical chip. (A) Electrical contact pads with the liquid flow cell and platinum reference and counter electrodes; (B) Higher magnification image showing the silicon nitride viewing window and glass carbon working electrode.

Macro Letters, (2015), http://dx.doi.org/10.1021/acsmacrolett.5b00479

Mag = 2.67 K X 1 μm WD = 4.9 mm EHT = 5.00 kV Signal A = InLens ESB Grid = 500 V Date :20 Oct 2011 Time :16:36:18 Auriga 60-39-95 H FIB Imaging = SEM Noise Reduction = Line Int. Done FIB Probe = 30KV:50pA System Vacuum = 1.38e-006 mbar

TEM Images of PEDOT Electrodeposition



Bright field TEM images of *in situ* electrochemical deposition of PEDOT. (A) Early stage in the process (right after initial CV deposition on bench) showing the dark PEDOT depositing from an aqueous solution onto a 20 μ m wide glassy carbon working electrode (anode) supported on a thin silicon nitride membrane. (B) Later stage in the process (after subsequent constant voltage deposition), showing the increased thickness and formation of rough protrusions at the edge of the PEDOT film. (C) Higher magnification view of the rough edge of the PEDOT film after further deposition.

Macro Letters, (2015), http://dx.doi.org/10.1021/acsmacrolett.5b00479

Mag = 2.67 K X 1 Auriga 60-39-95 占 EHT = 5.00 kV Signal A = InLens ESB Grid = 500 V Noise Reduction = Line Int. Done FIB Probe = 30KV:50pA

ESB Grid = 500 V Date :20 Oct 2011 Time :16 Probe = 30KV:50pA System Vacuum = 1.38e-000



Growth of PEDOT clusters during electrochemical deposition. (A) - (F) Captured frames from live video recording of in situ PEDOT deposition under TEM at constant voltage of +1.20 V; (G) Projected areas of individual PEDOT clusters measured from the video.

Auriga 60-39-95

WD = 4.9 mm

EHT = 5.00 kV

Noise Reduction = Line Int. Done FIB Probe = 30KV:50pA

Morphology of deposited PEDOT film



Optical and SEM images after electrochemical deposition. (A) Optical micrograph before deposition showing bare working electrode; (B) Optical micrograph after deposition showing accumulated dark PEDOT on the working electrode; (C) SEM image after deposition; (D) Higher magnification SEM image showing the "bumpy" surface morphology of the deposited PEDOT film.

Macro Letters, (2015), http://dx.doi.org/10.1021/acsmacrolett.5b00479

Mag = 2.627 X 1 µm Auriga 60-39-95

EHT = 5.00 kV

Noise Reduction = Line Int. Done FIB Probe = 30KV:50pA

Elemental mapping: EDS



EDS elemental mapping after in situ electrochemical deposition of PEDOT. (A) Secondary electron image; (B) sulfur (PEDOT); (C) carbon (PEDOT and glassy carbon working electrode); (D) chlorine (perchlorate dopant); (E) oxygen (PEDOT and perchlorate dopant) and (F) silicon (microfabricated substrate).

Auriga 60-39-95

EHT = 5.00 kVNoise Reduction = Line Int. Done FIB Probe = 30KV:50pA

Total dose ~ 0.1-1 mC/cm² Critical dose for beam damage $\sim 100 \text{ mC/cm}^2$ Features appear similar to those seen by SEM, optical

Next steps:

- Correlative Optical Microscopy
- Monomers
 - EDOT+, ProDOT, ProDOT+
- Solvents
- Counterions
- Fluid Flow Conditions
- Gels
- Living Cells

Macro Letters, (2015), http://dx.doi.org/10.1021/acsmacrolett.5b00479



EHT = 5.00 kV

Noise Reduction = Line Int. Done FIB Probe = 30KV:50pA

ESB Grid = 500 V

Date :20 Oct 2011 Time :16:36:18 System Vacuum = 1.38e-006 mba





1.tif Print Mag: 1690x @ 200mm 14:39 09/02/15 TEM Mode: Imaging

Auriga 60-39-95

10 microns HV=300kV Direct Mag: 200x X:Y: T: AMT Camera System

)ct 2011 Time :16:36:18 acuum = 1.38e-006 mbar



2.tif Print Mag: 1690x@200mm 14:45 09/02/15 Mag = 2.67 K X 1μm^{TEM Mode: Imaging}

Auriga 60-39-95

10 microns HV=300kV Direct Mag: 200x X:Y: T: AMT Camera System

ct 2011 Time :16:36:18 cuum = 1.38e-006 mbar



3.tif Print Mag: 1690x@200mm 14:53 09/02/15 TEM Mode: Imaging

Auriga 60<u>-39-95</u>

10 microns HV=300kV Direct Mag: 200x X:Y: T: AMT Camera System

2011 Time :16:36:18 ium = 1.38e-006 mbar



1.tif Print Mag: 1690x@ 200mm 14:39 09/02/15 TEM Mode: Imaging

r io intognig – ocia

10 microns HV=300kV Direct Mag: 200x X: Y: T: AMT Camera System

AMI Camera System Noise Reduction - Ellie III. Done - The Flobe - SonvisopA - Sjoten Pa

:t 2011 Time :16:36:18 cuum = 1.38e-006 <u>mbar</u>

Auriga 60-39-95

Galvanostatic Conditions 120 kV FEI Talos 200C, 150 nA



 Time - 60 s
 Time - 85 s
 Time - 105 s

 Mag = 2.67 K X
 1 μm
 WD = 4.9 mm
 EHT = 5.00 kV
 Signal A = InLens
 ESB Grid = 500 V
 Date :20 Oct 2011 Time :16:36:18

 Auriga 60-39-95
 Imaging = SEM
 Noise Reduction = Line Int. Done
 FIB Probe = 30KV:50pA
 System Vacuum = 1.38e-006 mbar

Potentiostatic Conditions Transmitted Light, +1.2 V, ~250 nA current



Initial droplets are translucent (PEDOT oligomers), liquid-like (breakup) Final film is dark (PEDOT), solid-like (stable)

Mag = 2.67 K X 1 μm WD = 4.9 mm EHT = 5.00 kV Signal A = InLens ESB Grid = 500 V Date :20 Oct 2011 Time :16:36:18 Auriga 60-39-95 H FIB Imaging = SEM Noise Reduction = Line Int. Done FIB Probe = 30KV:50pA System Vacuum = 1.38e-006 mbar t=210 sec

t=220 sec

t=230 sec

t=240 sec



Auriga 60-39-95



Time \rightarrow

Droplet Merging (M)

Droplet Breakup (B)

Droplet Evaporation (E)

Schematic diagram of the processes of droplet merging (M), droplet break-up (B), and droplet evaporation (E). These events are seen during imaging of liquid-like oligomer droplets formed during intermediate stages of the deposition, but are not seen in the solid, sessile PEDOT polymer films.

al A = InLens ESB Grid = 500 V Date :20 Oct 2011 Time :16:36:18 ne Int. Done FIB Probe = 30KV:50pA System Vacuum = 1.38e-006 mbar



Schematic of the hypothesized variations in the interaction of solid nanoparticles suspended in a flowing isotropic monomer solution with the products of an electrochemical polymerization reaction depositing on a solid surface. The initial liquid droplets form primarily from EDOT dimers that should have relatively weak interactions with the solid nanoparticles. The intermediate components (trimers and higher MW oligomers) will be viscoelastic and thus appear "sticky" to the particles, causing them to be trapped as they flow by, much as the mucus inside the nasal cavity and lungs traps unwanted contaminants. The final PEDOT polymer product will be solid, again resulting in relatively weak interactions that will allow the particles to continue to flow past. By examining the nanoparticle-containing solution with the electrodepositing film, we expect to obtain novel information about these complex rheological processes.

| Mag = 2.67 K X 1 | Auriga 60-39-95 | -

Nucleation and Growth Analysis



e :20 Oct 2011 Time :16:36:18 em Vacuum = 1.38e-006 mbar

In-situ Deposition of PEDOT: FEI Talos 200C 0.5 0.4 Droplet Density $(\#/\mu m^2)$ Droplet Area (µm²) 0.3 0.2 0.1 Time (secs) Auriga 60-39-95

Direct Observations of PEDOT / PDMICA Nanofibril Growth Mechanisms



PEDOT: thermochromic switching in external bias during CV



PEDOT-co-EDOT-acid

Auriga 60-39-95



20 µm 20 µm EHT = 5.00 kV ESB Grid = 500 V



Bright Field TEM t=0

Bright Field TEM t=300 secs of electrodeposition

Optical Microscopy t=300 secs of electrodeposition

ESB Grid = 500 V Date :20 Oct 2011 Time :16:36:18 FIB Probe = 30KV:50pA System Vacuum = 1.38e-006 mbar

EELS : UV/vis Spectroscopy



Figure 1. a) Raw EELS data and b) background-subtracted EELS data for rr-P3HT, PCBM, and PGeBTBT neat films. rr-P3HT films were annealed at 150 °C for 12 h to highlight crystalline properties. The background signal due to the zero-loss peak (<2 eV) is modeled as a third-order power law. EELS in (b) are generated by subtracting the background from the spectra in (a). EELS data are normalized by the maximum peak intensity.

Guo et al., "Probing Local Electronic Transitions in Organic Semiconductors through Energy-Loss Spectrum Imaging in the TEM", Adv. Funct. Mat., 25, 6071, (2015) DOI: 10.1002/adfm.201502090

Instrumentation limitations Beam sensitivity

Mag = 2.67 K X 1 μm WD = 4.9 mm EHT = 5.00 kV Signal A = InLens ESB Grid = 500 V Date :20 Oct 2011 Time :16:36:18 Auriga 60-39-95 H FIB Imaging = SEM Noise Reduction = Line Int. Done FIB Probe = 30KV:50pA System Vacuum = 1.38e-006 mbar

Microfocused Raman



Nano-IR and Nano-Raman



Anasys NanoIR2



- Expands nanoscale IR to a broad range of real world samples
- New resonance enhanced mode enables nanoscale IR on <20nm films
- Rich, interpretable IR spectra
- Powerful, full featured AFM
- Multifunctional measurements including integrated thermal and mechanical property mapping
- Designed and built for productivity and rapid time-to-results



Auriga 60-39-95

EHT = 5.00 kV

New Materials Design

Thiol-ene "click" chemistry:

- POSS-ProDOT cross-linkers
- Alternating Copolymers
- Conjugated Polymer Networks

EDOT-amine: adhesion, transport





Ouyang, et al., **Science Advances**, 3: e1600448, (2017).

Bin Wei

Functionalized EDOT monomers



Samadhan Nagane, Peter Sitarik, Yuhang Wu, Quintin Baugh, Shrirang Chhatre, Junghyun Lee, and David C. Martin, "Functionalized Polythiophene Copolymers for Electronic Biomedical Devices", MRS Advances, 1-14, (2020). http://dx.doi.org/10.1557/adv.2020.3



Auriga 60-39-95

EHT = 5.00 kV

Noise Reduction = Line Int. Done FIB Probe = 30KV:50pA